

# **Nodal flow and** *situs inversus***: A review of the literature**

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#### **ABSTRACT**

Variations of situs solitus totalis, the normal organ disposition pattern, involve dextrocardia, situs inversus, situs inversus totalis and situs ambiguous. These laterality defects have been documented for more than 400 years, and a connection to ciliary abnormalities has been elucidated by observing cilia from the primitive node, which can promote "nodal flow", possibly necessary for the development of left-right asymmetry. The aim of

#### **LIST OF ABBREVIATIONS AND ACRONYMS**

Cerl2 - Cerberus-like 2 FGF8 - Fibroblast Growth Factor 8 LPM-Mesoderma Get Side Plate NVP - Parcela vesicular nodal PCP - Planar Cell Polarity SHH - Sonic Hedgehog RA - Retinoic Acid TGF-β - Transforming Growth Factor Beta TIF - Intraflagellar Transport

this work was to perform a literature review, showing variations in the usual anatomy and the connection between nodal flow and normal asymmetric pattern. A narrative review was carried out with articles from the MEDLINE/PubMed, SciELO and LILACS databases and with books relevant to the theme. The initial embryonic development comprises the formation of the organizing structure, a primitive node, and its cilia are responsible for the nodal flow, determinant in many vertebrates in the breaking of the embryonic bilateral symmetry. The formation of asymmetry is associated with morphogenic gradients to the left of the node, the model of two cilia and the mechanism of nodal vesicular plots, until it results in organogenesis. Ciliary abnormalities configure clinical manifestations in disorders such as Primary Ciliary Dyskinesia and Kartagener syndrome. The nodal flow, in fact, is important in normal anatomical standardization, and the cilia are necessary for the configuration of situs solitus totalis.

**Keywords:** Nodal flow, Situs Inversus, Laterality defects, Primary Ciliary Dyskinesia, Kartagener syndrome.

#### **1 INTRODUCTION**

The normal anatomical pattern, with all structures in their usual position, is termed *situs solitus totalis*. There are variations of this standardization, which may or may not be associated with important functional changes: *situs solitus* with dextrocardia, in which the abnormality is the presence of the



heart apex on the right side; *Inversus Sites*, with the organs located in an inverted position, but without dextrocardia; *situs inversus totalis*, with all organs reversed, and *ambiguous site*, or heterotaxy, in which the thoracoabdominal organs do not follow a normal pattern of arrangement, may be partially inverted, arranged in the midline, or symmetrical bilaterally. Heterotaxy, which often causes serious health problems, is divided into two categories: *ambiguous site* with polysplenia, or left isomerism, with a variable number of bilaterally bilobed spleens and lungs, and *ambiguous site* with asplenia, or right isomerism, with absence of spleen and bilaterally trilobed lungs. Patients with these changes are likely to have other malformations, especially heart defects  $1-3$ .

In the general anatomical context, considering in addition to variations in cardiac morphology, known laterality defects, characterized by the abnormal arrangement of structures and organs, have been documented for more than 400 years, with many reports, however, underlying causes unknown. Girolamo Fabrizio was the first to describe such abnormalities, around 1600, followed by Marco Aurelio Severino, who in 1643 documented a case of human dextrocardia, and by Matthew Baillie, responsible for describing a complete reversal of the thoracoabdominal organs in 1788<sup>4,5</sup>.

In 1995, Afzelius' published study demonstrated a connection between changes in ciliary structures and the *Inversus Sites*, one of the laterality defects of the general anatomy <sup>6</sup>. This correlation can be elucidated by the evidence of studies on the gastrulation of mouse embryos, which demonstrated that the primitive node, characterized as an organizing structure, is made up of single-cilium presenting cells, that is, each cell has a monocilium  $<sup>7</sup>$ .</sup>

In the fully developed primitive knot, there are a total of 200 to 300 cilia, each of these measuring 7 to 10 micrometers in length and spaced at intervals of 5 to 10 micrometers. Central nodal cells contain monocilia that exhibit motility, while peripheral nodal cells (crown cells) exhibit immobile monocilia. The central cilia, in such a way, are able to produce a rotation and generate a directed flow of extra-embryonic fluid to the left through the node. This fact supports the existence of a "nodal flow" model, which would be necessary for the development of the left-right asymmetry of the embryo and, therefore, for a normal arrangement of anatomical structures  $^{7, 8}$ .

Knowledge about the mechanisms involved in the determination of physiological left-right asymmetry is essential for understanding the causality of various laterality defects. The study of these abnormal anatomical states in characteristic disorders or in isolation allows us to elucidate not only the triggering factor of the problem, but also to establish the correlation between the embryonic abnormality involved and the clinical one presented. From this perspective, the aim of this study is to present a review of the existing literature in this sphere, highlighting the variations *of the habitual situs* in a general anatomical context – without limiting to the nomeclatures of the cardiac context – and the link between the known nodal flow and the standardization of normal asymmetry.



### **2 METHODOLOGY**

The present work consists of a narrative review of the literature, based on scientific articles selected from the MEDLINE/PubMed, SciELO (Scientific Electronic Library Online) and LILACS (Latin American and Caribbean Health Sciences Literature) databases, as well as important books for the explanation of the theme. Pertinent data and information were collected, with the search for studies using descriptors related to the subjects addressed and without a pre-established publication period.

#### **3 DEVELOPMENT**

### 3.1 EARLY EMBRYONIC DEVELOPMENT

During the first week of human development, in the blastogenesis stage, the conceptus – formed by the embryo and its membranes – gives rise to the blastocyst. In this study, the separation of blastomeres by fluid in the blastocystic cavity configures the trophoblast and the embryoblast, which constitute, respectively, the formation of its wall and a projection into the blastocystic cavity. With the gradual degeneration of the zona pellucida, the blastocyst grows rapidly and is adhered to the endometrial epithelium. From this moment on, the trophoblast proliferates and differentiates into two layers: the cytotrophoblast (inner layer) and the syncytiotrophoblast (outer layer) which, with its enzymatic synthesis, enables the effective implantation of the blastocyst in the endometrium  $9,10$ .

As the progress of blastocyst implantation continues, in the second week of development, morphological changes in the embryoblast, responsible for the formation of the embryo, lead to the generation of the bilaminar embryonic disc, consisting of the epiblast and the hypoblast. The epiblast contributes to the formation of the amniotic cavity, and the hypoblast to that of the exocoelomic cavity. The formation of the bilaminar disc in this way defines the primitive dorsoventral axis of the embryo. In the third week of development, the bilaminar embryonic disc finally converts into a trilaminar embryonic disc, characterizing the gastrulation event. The three germ layers (embryonic ectoderm, endoderm and mesoderm) formed during the stage give rise to specific organs and tissues, constituting the principle of morphogenesis  $^{7, 9, 11}$ .

Gastrulation begins with the formation of a longitudinal structure in the midline of the epiblast surface: the primitive line, close to the caudal region of the embryonic disc, resulting from an induction of epiblastic cells by the extraembryonic region. This formation defines the main body axes: craniocaudal, mediolateral, dorsoventral, and left-right axes. The cephalic end of the primitive line forms the primitive node, considered an organizing structure; It contains a circular depression called the primitive fosseta, which is continuous caudally with the primitive sulcus, a depression developed in the line. The pit and sulcus are the result of an invagination of cells from the primitive line. Some of these cells, which migrate to the interior of the embryonic disc, invade the hypoblast, forming a layer of definitive endoderm. Others migrate bilaterally, between the endoderm and the epiblast, to



form the intraembryonic mesoderm. The remaining epiblastic cells form the embryonic ectoderm  $10$ , 12 .

# 3.2 CILIARY STRUCTURE

Cilia can be defined as slender extensions that protrude from the surface of different cells. They consist of a cytosolic axis – the ciliary matrix – surrounded by an extension of the plasma membrane; Inside, this matrix comprises the axoneme, a filamentous scaffold that follows the longitudinal axis of the ciliary structure and is formed by microtubules. These microtubules are made up of protofilaments and sprout from the basal corpuscle (or basal body), which is located inside the cell, under the plasma membrane, and anchors the cilia. In addition, they have a positive end and a negative end, the latter facing the corpuscle. The axoneme and basal body have doubles (doublets) and trios of microtubules, respectively, and are connected by a transition zone. The A and B microtubules of the axoneme doubles are continuous with the A and B microtubules of the basal corpuscle cracks  $^{13, 14}$ .

The processes of formation and maintenance of cilia and their counterparts, the flagella, occur through the intraflagellar transport system (TIF), with motor proteins associated with microtubules kinesin and cytoplasmic dynein - involved. Under a different system, the fundamental process for the movement of the structures requires a motor protein of the distinct dynein class: axonemal dynein <sup>15</sup>, whose motive activity, like that of TIF proteins, also requires the hydrolysis of ATP as an energy source <sup>16</sup>. Axonemal dynein contains internal and external arms that protrude from the A microtubule of the axoneme doubletes and interact with the B microtubule of adjacent doublets, forming dynein bridges; through its ATPase activity, then, the portions of the protein associated with the B microtubule slide along the microtubule toward the negative end. As a result, motor proteins promote the sliding of one doublet in relation to the other and induce ciliary curvature, thus characterizing the dynamics of cilia and flagella beating <sup>13, 17</sup>.

There are three basic categories of eyelashes: motile, primary, and nodal. The motile cilia, commonly found in epithelia responsible for transporting secretions, and the flagella, considerably longer and single by cells, have a  $9 + 2$  axonemal organization, that is, they have 9 microtubule doublets surrounding 2 central microtubules; Microtubules are associated with motor proteins necessary for ciliary motility. Primary cilia are immobile cellular projections in an arrangement of 9+0 microtubules (9 microtubule doublets, without a central pair), with an absence of associated motor proteins. They are found in cells such as those of the renal ducts and in the epithelium of the bile ducts, and curve passively as a result of the flow of fluids; Thus, they are able to act as sensory antennas and to generate and transmit signals to the intracellular medium in response to extracellular events. In addition, the primary cilia are also involved in the transduction of the Hedgehog signal active in the early development of vertebrates. Finally, the nodal cilia, found in the embryo, have an axonemal



organization similar to that of the primary cilia; however, they have associated motor proteins, having the ability to perform active movement. The absence of the central pairs of microtubules is possibly responsible for their rotational movement in a full cone trajectory, in contrast to the  $9 + 2$  motile cilia, whose trajectory is in a half cone  $^{18, 19}$ .

The nodal cilia with  $9 + 0$  axonemal architecture, then, are responsible for the dynamics of the nodal flow, which occurs at a speed of 15 to 20 μm/s, specifically between the stages of one to two somites and the six-somite stage  $20$ . Still, the rotational movement is established in a clockwise direction, unlike the eyelashes commonly found  $2<sup>1</sup>$ . Studies in vertebrates also demonstrate a posterior positioning of these cilia at a time coinciding with the onset of the flow, specifically to the left. Initially, they are projected into the center of the cells, but apparently become posterior by movement from the basal body to the posterior cell pole <sup>22</sup>. This positioning of monocilia is determined by the signaling of the PCP (Planar Cell Polarity) pathway in some vertebrates  $^{23}$ .

In addition, the dominant unidirectional character of the flow to the left is made possible by the posterior inclination of these cilia from a vertical angle, with the trajectory of their tip displaced posteriorly when compared to their root; With this configuration, the cilia swing to the left, away from the surface of the cell, and move to the right, towards the surface of the cell. In this way, there is a movement of fluid directed specifically to the left, since, according to hydrodynamics, the surface of the cell is stationary and slows down the movement of fluid by shear resistance and, as an effect, the sweep to the right is less effective than the movement to the left  $^{22, 24}$ .

# 3.3 DETERMINATION OF THE LEFT-RIGHT PLANE

In all vertebrates, despite the perceptible external symmetry between the right and left sides, there are processes that trigger an asymmetrical pattern of organization of internal structures and organs, as well as of the vascular network <sup>25</sup>. The fundamental processes to generate asymmetry require different mechanisms. Initially, a mechanism is needed to break the bilateral symmetry of the embryo, such as a molecular asymmetry. Subsequently, signaling of this small-scale generated asymmetry must be established in larger regions of the embryo and ultimately result in an asymmetric organogenesis. In this way, the normal development of the left-right configuration results in the so-called *situs solitus*  <sup>8</sup>. The left-right axis is possibly determined in relation to the anteroposterior and dorsoventral axes, these being the axes along which embryonic morphogenesis occurs  $^{26}$ .

The onset of the asymmetric left-right configuration in vertebrates is a controversial process. In mice, however, experiments suggest that symmetry breaking begins with the generation of extraembryonic fluid flow to the left: the well-known nodal flow, which occurs through the primitive node  $20$ . The node is described as a transient structure of the midline, with a "ciliated well" on the ventral surface, which is important for the determination of left-right asymmetry. This activity can be proven



by the role of the monocilia of the nodal cells, which with their rotational movements, are responsible for generating this nodal flow. Analyses of experiments with fluorescent beads added to the liquid in the node region demonstrated the activity of cilia in the nodal flow: in normal wild embryos, these beads moved unidirectionally to the left, but in mutant embryos with absence of the motor proteins KIF3A and KIF3B, belonging to the kinesin superfamily, they showed Brownian motion (random)  $^{24}$ .

The application of artificial flux in cultured mouse embryos also evidenced the relationship between nodal flux and the beginning of the asymmetry process. It was observed that a rapid flow directed to the right resulted in a reversal of the normal flow to the left and a reverse left-right development. On the other hand, the application of a left-directed flow in embryos with mutations affecting inversin – a protein with ankyrin repeats –, which had slow nodal flow, and left-to-right axonemal dynein, which had monociliary paralysis, was sufficient to recover the normal asymmetric pattern. Several mutations are related to the impairment of left-right development, and some of the genes involved are responsible for the formation of the embryonic knot itself<sup>8</sup>.

Hirokawa *et al* <sup>24</sup> considered an absence of an asymmetric event prior to nodal flow in mice. Thus, they discuss the possibility that flow has become the main mechanism of asymmetry with the emergence of viviparous animals and that it is sufficient to determine laterality in mammals. Dasgupta and Amack  $^{22}$  report that many vertebrate embryos have hair structures analogous to the primitive mouse knot that are responsible for an asymmetric flow, such as Kupffer's vessicle in certain fish, the gastrocele teat plate in the frog, and the posterior notochordal plate in the rabbit, characterizing leftright organizers. However, they point out that cilia would not be necessary to generate asymmetry in all vertebrates. Hensen's knot, as exemplified, holds the first asymmetric gene expression in the chick, however, this structure may not present motile cilia, since mutants with defective cilia present normal development of asymmetry. In addition, they report that the knot of the pig embryo does not have cilia, as well as is not exposed to an extra-embryonic liquid – a factor also present in the cow embryo – unlike what occurs in the mouse embryo. Thus, they demonstrate that some vertebrates, including mammals, can use mechanisms independent of cilia in the development of asymmetry, unlike the idea that nodal flow, *for himself*, is sufficient.

Considering an asymmetric information generation based on the mechanism of nodal flow, this can be elucidated, in principle, from two proposed models. The first concerns the formation of a gradient of morphogens on the left side of the node – equivalent to the mouse's left-right organizer; The chemical morphogens would be secreted into the node and transported to the left through the nodal flow. The second proposes the existence of a physical stimulation generated by the flow, which would be mechanically detected by the immobile cilia of the peripheral region of the node, being called the "two-eyelash model", since there are distinct cilia responsible for both the generation of the flow and its perception  $24$ .



Mc and Brueckner <sup>8</sup> clarify that, in fact, the flow may be responsible for generating a morphogenic gradient on the left side of the node; The morphogenic molecule would then be able to configure an asymmetric gene expression cascade on the left. Many molecules found in the structure, such as Nodal, perform an activity in left-right development, and are therefore candidates for the role of morphogen. Genes expressed on a single side of the embryonic midline employ mechanisms to propagate signals between cell subpopulations, and this occasionally results in asymmetric morphogenesis of specific structures. In general, the gene products responsible for asymmetry are not only involved in this process, and some of them are not even present asymmetrically  $^{26}$ .

Nodal is a signaling molecule belonging to the transforming growth factor beta (TGF-β) family of proteins, initially expressed bilaterally in the crown cells of the mouse organizer. The protein produced in the node can migrate to the left side plate mesoderm (MPL) and thus activate the expression of the gene itself *Nodal*; the asymmetric manifestation in the LPM begins at the two-somite stage and ends at the six-somite stage, leading to the conclusion that nodal flow may occur specifically to initiate expression *Nodal* on the left-hand side. The cells of the left LPM that received the Nodal Signaling then contribute to the specific morphological features of the left side, while those of the right side, which did not, contribute to the characteristics of the right  $20, 27$ .

Fibroblast growth factor 8 (FGF8) transported by nodal flow also seems necessary for left-right determination by some mechanism, since mice deficient in *Fgf8* have an absence of expression *Nodal* in LPM. In mammals, Nodal signaling also induces the expression of Lefty genes: *Lefty1*, predominantly in the midline of the embryo, and *Lefty2*, predominantly in the left MPL. The absence of any of these – *Lefty1*, which acts as a barrier on the midline, or *Lefty2*, which acts as a Nodal feedback inhibitor – leads to a "leakage" of the mesoderm Nodal signal from the left lateral plate to the right side, resulting in expression *Nodal* bilateral and alteration of normal morphology <sup>20, 26</sup>.

Beyond Genes *Nodal* and *Lefty2*, *Pitx2* it is also expressed in the left LPM, and its asymmetric expression is induced by nodal signaling. The transcription factor Pitx2 is considered the main regulator of asymmetric organogenesis at the molecular level, but there are mechanisms independent of the factor also operating. Nodal activity, which is necessary for the cascade of asymmetric expressions, can also be controlled by a protein belonging to the TGF-β antagonist family, Cerl2 (Cerberus-like 2). At first, the protein is found in crown cells on either side of the node, in the early stage of mouse development. Cerl2 then accumulates on the right side and prevents the Nodal from acting on the right LPM. Subsequently, the protein is translocated by the nodal flow to the left, terminating nodal activity on the left side of the node and in the left LPM at a precise time  $20, 28$ .

On the other hand, in the "two-eyelash model", the distinct characteristics of the mouse organizer's monocilia may provide a basis for the hypothesis. Monocilia that are motile, centrally located, contain the motor protein Lrd (left-right dynein) and the protein polycystin-2, a cation-



permeable channel; the immobile cilia, located peripherally, lack the Lrd protein, but have polycystin-2. This protein is a calcium-activated channel, which in the primary cilia of the renal epithelium functions as a mechanotransducer, increasing the concentration of intracellular calcium in response to fluid flow. In the nodal structure, it is evident that, while the Lrd-bearing monocilia generate the nodal flow, the immobile monocilia, which do not have the Lrd protein, can function as mechanosensory cilia. Thus, they detect nodal flow to the left, and the existing polycystin-2 leads to an increase in intracellular calcium on the left side of the node, generating asymmetric signaling. The calcium signal resulting from mechanosensitivity can then trigger the expression of a specific growth factor to the left of the node 25, 29 .

A third mechanism proposed for the generation of asymmetric information by nodal flow – analogous to the first model – concerns the nodal vesicular parcels (NVPs). NVPs are materials secreted from the surface of the mouse node that are transported to the left through the flow. These vesicles are made up of lipoprotein particles coated by a membrane; They may also comprise a content of signaling molecules, such as Sonic Hedgehog (SHH), Retinoic Acid (RA) and other morphogens. NVPs appear to fragment on the left side of the node, with the help of peripheral cilia, through the interaction of the SHH of the vesicles with its receptor called Smoothened in the cilia, and are absorbed by the nodal surface. Thus, they may be responsible for producing a concentration gradient to the left of the node, and the molecules released may play a role in the left-right determination. In addition, they may also be responsible for an asymmetric elevation of intracellular calcium through Hedgehog signaling  $21, 24$ .

Shiratori and Hamada  $^{20}$ , however, expose the fact that SHH plays an important role in leftright determination in avian species, but in distinct vertebrates, such as the mouse, it does not seem to be directly involved in asymmetric determination, but rather in the formation of a functional midline. Similarly, AR would be related to the maintenance of bilateral symmetry during the formation of somites, but not to the left-right determination itself. With this, the authors refute the potential role of NVP content in the de facto establishment of the asymmetric configuration in other vertebrates.

Macroscopically, at least three distinct mechanisms are involved in the formation of anatomically asymmetric structures: the first concerns the directional "looping" of initially tubular organs, such as the heart, which go through a series of steps (looping, flexion and rotation) and reach their correct final positioning; the second corresponds to differential branching, in which a pair of symmetrically formed organs, on both sides, acquires differences in their size or branching pattern, as occurs with the lungs and; The third – unilateral regression – is related to the disappearance of one side of a symmetrical structure, such as a blood vessel  $^{20}$ . Failure in the processes of establishing adequate left-right asymmetry results in a range of laterality defects, such as absence of breaking bilateral



symmetry, independent localization of structures in relation to the *Site* normal and partially or totally abnormal asymmetric orientation  $26, 29$ .

# 3.4 CILIARY ABNORMALITY

Alterations in genes associated with cilia can lead to changes in their structure and affect their signaling mechanisms in various anatomical locations. The gene *PKHD1* (Polycystic Kidney and Hepatic Disease 1), for example, encodes the protein fibrocystin, or polyductin, expressed in the axoneme and basal body of primary cilia. Mutations in the gene lead to autosomal recessive polycystic kidney disease, a severe form of pediatric cystic kidney disease <sup>30</sup> . Genes *PKD1* (Polycystic Kidney Disease 1) e *PKD2* (Polycystic Kidney Disease 2) encode polycystin proteins, which are also located in the primary cilia and play a necessary role in renal function. Mutations involving these genes are responsible for the ciliopathy of autosomal dominant polycystic kidney disease, or adult polycystic kidney disease 31, 32.

Ciliary genes *BBS* (Bardet-Biedl Syndrome) Mutants cause Bardet-Biedl syndrome, a rare autosomal recessive disorder that leads to varied manifestations such as obesity, renal abnormalities, hypogonadism, and retinopathy <sup>33</sup>. It is also characterized by involvement of the primary cilia, since the BBS proteins normally encoded are located in the basal body and in the ciliary axoneme and contribute to the formation of an active complex in the intraflagellar transport system  $34$ .

Other mutations that lead to the absence or immotility of the cilia can lead to abnormalities in the left-right organization, confirming their essential role in generating the directed nodal flow and, consequently, the initial process of symmetry breaking. The mice with the gene *Dvl* mutant – the gene responsible for encoding Dishevelled, the cytoplasmic protein of the PCP pathway – present interruption of the posterior positioning of the basal bodies and randomization of the flow, which also determines a defective left-right standardization <sup>35</sup>.

# **3.4.1 Changes in the handedness of the heart**

Cardiac anatomy also involves the characterization of *Situs* and its complexities. Or *situs solitus*  The atrial lesion corresponds to the usual arrangement of the morphologically right atrium to the right and the left to the left, while the mirror image of this situation corresponds to the *Inversus Sites*. The presence of atrial isomerism, with both atria presenting the right or left morphology, constitutes the *Situs ambiguous*. In addition, the atrioventricular connection is characterized as concordant, when the right atrium is connected to the right ventricle and the left atrium to the left ventricle, and discordant, when the right atrium is connected to the left ventricle and the left atrium to the right ventricle; When the atria are connected to a single ventricular chamber, the univentricular connection is configured. The venoatrial connection is also analyzed, observing how the systemic and pulmonary veins are



connected to the atrial chambers. Finally, the ventriculoarterial connection is classified as: concordant, when the aorta emerges from the left ventricle and the pulmonary trunk from the right ventricle; discordant, when the opposite occurs; double outflow tract, when the two vessels emerge from the same ventricle; and single outflow tract, when only one arterial vessel emerges from the heart or there is aortic or pulmonary atresia 36, 37.

# 3.5 IN THE MEDICAL CLINIC

# **3.5.1 Primary Ciliary Dyskinesia**

Primary ciliary dyskinesia (PCD) is a classically autosomal recessive disorder, rarely linked to the X chromosome, caused by mutations in more than 40 genes responsible for coding proteins necessary for the proper assembly and functioning of motile cilia. Different mutant genes determine different symptomatology severities, which depend on the extent of ciliary structural and functional alterations. Mutations in the gene *DNAH5* (Dynein Axonemal Heavy Chain 5), for example, which encodes the outer arm heavy chain of dynein from the proximal and distal zones of the cilium, lead to ciliary immotility, while mutations in the gene *DNAH9* (Dynein Axonemal Heavy Chain 9), which encodes the heavy chain of the outer arm of dynein from the distal zone, leads only to a reduced curvature of the distal ciliary portion and does not alter the beating rate  $38,39$ .

The disorder is characterized by generalized ciliary dysfunction and, therefore, there are ciliary impairments in distinct anatomical regions, such as the epithelium of the respiratory tract, the epithelium of the vas deferens, the fallopian tubes, and the ependyma, as well as sperm involvement. Thus, the disease predisposes to dysfunctional mucociliary clearance and recurrent respiratory infections, and it is important to consider it as a differential diagnosis in cases of chronic infections of the respiratory system, and it is also frequently associated with infertility and hydrocephalus, in addition to other clinical manifestations (Table 1). The presence of other clinical indicators also raises the suspicion of PCD, including a family or personal history of ciliopathies and the existence of laterality disorders, since motility cilia play an essential role in the left-right pattern  $40, 41$ .

1. ГЕНЕ 1 - СПИСАН ПІАННСЬКАНОПУ ОТ РИННАГУ СИНАГУ СРУБЕНІСЬКА	
Lung	Respiratory distress syndrome, pneumonia,
	atelectasis (neonatal period)
	Chronic productive cough
	<b>Bronchorhoea</b>
	Recurrent episodes of pneumonia
	Severe and/or atypical "bronchial asthma"
	unresponsive to conventional therapy
	<b>Bronchiectasis</b>
	Digital Hypocratism
Middle ear	Chronic serous otitis media
	Transmission hearing loss
	Persistent otorrhea after tympanostomy

TABLE 1 - Clinical manifestations of primary ciliary dyskinesia







The definitive diagnosis is established when at least three of the following phenotypic manifestations coexist: respiratory distress in full-term infants, chronic nasal congestion and chronic productive cough throughout the year, chronic otitis media with effusion for more than 6 months, chronic pansinusitis, bronchiectasis and other recurrent lower respiratory tract infections, and finally male infertility, laterality defects and family history of the disease. When there is a concomitant presence of bronchiectasis, pansinusitis, and *situs inversus totalis*, one of the existing laterality disorders, the diagnosis is Kartagener's syndrome <sup>41, 42</sup>.

# **3.5.2 Kartagener syndrome**

Kartagener syndrome is a subgroup of primary ciliary dyskinesia, being the most severe clinical form of the disorder and being present in 50% of cases. It is related to the deficiency of ciliary dynein, protein Responsible for the generation of mechanical force in the movement of the eyelashes. It is also known as Immobile Eyelash syndrome, associated with the absence of frontal sinuses, chronic rhinosinusitis, and bronchiectasis, with recurrent respiratory infections and airway damage. Spermatozoa are immobile, and the male individual is a carrier, infertile  $40, 43$ .

The main picture of a child born with the disorder is based on pulmonary symptoms, which become evident within 24 hours post-birth and which, in a significant number of cases, cause neonatal respiratory distress syndrome. Other typical symptoms associated with seeking medical help refer to recurrent ear infections, persistent wet cough, nasal congestion, and chronic wheezing. In accordance with the abnormalities that accompany PCD, Kartagener syndrome also presents conditions justified by ciliary alterations, which lead to abnormal positioning of certain organs and alterations in the



function of other structures. Approximately 20% of patients with *Inversus Sites* belong to the group with Kartagener syndrome <sup>44, 45</sup>.

# **3.5.3** *Inversus Sites*

Or *Inversus Sites* It is characterized as a mirror-image arrangement of the abdominal organs, but with a *Situs* normal cardiac disease, i.e. the apex is located in the left hemithorax and therefore there is a levocardia. On the other hand, the *situs inversus totalis* It is characterized by the mirror image of the *situs solitus totalis*, including reverse-located abdominal viscera, right lung bilobed and left lung trilobed and dextrocardia. Thus, the individual exhibits completely inverted left-right asymmetry, with transposition of the viscera in the thorax and abdomen  $46, 47$ . The abnormality is typically asymptomatic and is not considered a premalignant condition, however, a small number of cases of cancer have been reported, as well as cardiovascular malformations, bowel anomalies, and respiratory comorbidities <sup>48</sup>.

This abnormality is inherited as an autosomal recessive genetic trait and can occur in combination with primary ciliary dyskinesia. However, some genes are responsible for the occurrence of *Inversus Sites* and are not associated with the occurrence of PCD, and the mechanisms involved in this isolated laterality alteration are based on the role of some of these genes in the coding of ciliary proteins. Mutations in the gene *CFAP52* (Cilia and Flagella Associated Protein 52), for example, which encodes a protein active in ciliary signal transduction processes, establish causality with the occurrence of the *Inversus Sites* and heterotaxy in patients who do not have dyskinesia<sup>49, 50</sup>.

In the United States, an average distribution of 1 per 10,000 individuals has been observed, however, the discovery of such morphological alteration does not depend on symptoms resulting from the inversion of the organs themselves, often being discovered in the investigation of unrelated health alterations, or even at the time of necropsy. This can be observed with the American patient Rose Marie Bentley, who died of natural causes, but as soon as her body was donated for research to a university in Portland, Oregon, the students of an anatomy class noticed that many of her organs were mirrored 51 .

# **4 FINAL THOUGHTS**

The scientific evidence based on the analyzed studies denotes the role that, in fact, the nodal flow can play in the establishment of the normal left-right asymmetry in certain vertebrates and, therefore, in the usual anatomical pattern. In addition, the clinical presentation by patients with ciliary disorders reaffirms the conception that eyelashes are involved in the definition of a pattern of situs *solitus totalis*. As a result, the laterality defects that initially had an undefined causality can be progressively understood and, thus, the need for a constant acquisition of knowledge in the field of



embryology is noted, aiming to elucidate the various abnormalities that involve human development and the mechanisms involved in their occurrence.



# **REFERENCES**

MUJO, T.. FINNEGAN, T.. JOSHI, J.. WILCOXEN, K. A.. REED, J. C. Situs ambiguous, levocardia, right sided stomach, obstructing duodenal web, and intestinal nonrotation: a case report. J Radiol Case Rep. 2015;9(2). p. 16-23.

JUNCOS C., María. ROS F., María Amparo. MARAVALL LL, María. ÁLVAREZ-PITTI, Julio. Situs inversus totalis: a propósito de 2 casos clínicos. Rev chil pediatr. 2014;85(3). p. 344-50,.

LAGROTTA, G.. MOISES, M. Heterotaxy Polysplenia Syndrome in Adulthood: Focused Review and a Case Report. Cureus. 2020;12(1). p. e6822.

CHANDRARAJ, S. Observations on some additional abnormalities in situs inversus viscerum. J Anat. 1976;122(Pt 2). p. 377-88.

PENNEKAMP, P.. MENCHEN, T.. DWORNICZAK, B.. HAMADA, H. Situs inversus and ciliary abnormalities: 20 years later, what is the connection? Cilia. 2015;4(1). p. 1.

AFZELIUS, B. A. Situs inversus and ciliary abnormalities. What is the connection? Int J Dev Biol. 1995;39(5). p. 839-44.

SCHOENWOLF, S.. BLEYL, S.B.. BRAUER, P.R.. FRANCIS-WEST, P.H. Larsen: Embriologia Humana. 5ª ed. Rio de Janeiro: Elsevier; 2015. 576 p.

MCGRATH, J.. BRUECKNER, M. Cilia are at the heart of vertebrate left-right asymmetry. Curr Opin Genet Dev. 2003;13(4). p. 385-92.

MOORE, K.L.. PERSAUD, T.V.N.. TORCHIA, M.G. Embriologia Clinica. 10ª ed ed. Rio de Janeiro: Elsevier; 2016. 552 p.

SADLER, T. W. Langman: Embriologia médica. 13ª ed ed. Rio de Janeiro: Guanabara Koogan; 2016. 348 p.

CARLSON, B.M. Embriologia Humana e Biologia do Desenvolvimento: Elsevier Editora Ltda.; 2014. 520 p.

STANDRING, S. Gray's anatomia: A base anatômica da prática clínica. 40ª ed ed. Rio de Janeiro: Elsevier Editora Ltda.; 2010. 1583 p.

DE ROBERTIS, E. M.. HIB, J. Biologia celular e molecular. Rio de Janeiro: Guanabara Koogan; 2014.

ISHIKAWA, T. Axoneme Structure from Motile Cilia. Cold Spring Harb Perspect Biol. 2017;9(1). p.

KIERSZENBAUM, A.L.. TRES, L. L. Histologia e biologia celular: uma introdução à patologia. 4 ed. ed. Rio de Janeiro: GEN Guanabara Koogan; 2016. 752 p.

NELSON, D.L.. LEHNINGER, A.L.. COX, M.M.. FOIX, C.M.C.. LEÓN, S.. ROCA, J.V. Lehninger principios de bioquímica. 6 ed ed. Porto Alegre: Artmed; 2013. 1328 p.

LODISH, Harvey. BERK, Arnold. KAISER, Chris A.. KRIEGER, Monty. BRETSCHER, Anthony. PLOEGH, Hidde. Biologia celular e molecular. 7<sup>ª</sup> ed. ed. Porto Alegre: Artmed; 2014. 1244 p.



PAWLINA, W. Ross Histologia: Texto e Atlas. 7º ed ed. Rio de Janeiro: Guanabara Koogan; 2016. 984 p.

GOETZ, S. C.. ANDERSON, K. V. The primary cilium: a signalling centre during vertebrate development. Nat Rev Genet. 2010;11(5). p. 331-44.

SHIRATORI, H.. HAMADA, H. The left-right axis in the mouse: from origin to morphology. Development. 2006;133(11). p. 2095-104.

HIROKAWA, N.. TANAKA, Y.. OKADA, Y. Left-right determination: involvement of molecular motor KIF3, cilia, and nodal flow. Cold Spring Harb Perspect Biol. 2009;1(1). p. a000802.

DASGUPTA, A.. AMACK, J. D. Cilia in vertebrate left-right patterning. Philos Trans R Soc Lond B Biol Sci. 2016;371(1710). p.

WALLINGFORD, J. B.. MITCHELL, B. Strange as it may seem: the many links between Wnt signaling, planar cell polarity, and cilia. Genes Dev. 2011;25(3). p. 201-13.

HIROKAWA, N.. TANAKA, Y.. OKADA, Y.. TAKEDA, S. Nodal flow and the generation of leftright asymmetry. Cell. 2006;125(1). p. 33-45.

NORRIS, D. Breaking the left-right axis: do nodal parcels pass a signal to the left? Bioessays. 2005;27(10). p. 991-4.

LEVIN, M. Left-right asymmetry in embryonic development: a comprehensive review. Mech Dev. 2005;122(1). p. 3-25.

NAKAMURA, T.. MINE, N.. NAKAGUCHI, E.. MOCHIZUKI, A.. YAMAMOTO, M.. YASHIRO, K., et al. Generation of robust left-right asymmetry in the mouse embryo requires a self-enhancement and lateral-inhibition system. Dev Cell. 2006;11(4). p. 495-504.

INACIO, J. M.. MARQUES, S.. NAKAMURA, T.. SHINOHARA, K.. MENO, C.. HAMADA, H., et al. The dynamic right-to-left translocation of Cerl2 is involved in the regulation and termination of Nodal activity in the mouse node. PLoS One. 2013;8(3). p. e60406.

MCGRATH, J.. SOMLO, S.. MAKOVA, S.. TIAN, X.. BRUECKNER, M. Two populations of node monocilia initiate left-right asymmetry in the mouse. Cell. 2003;114(1). p. 61-73.

DIAS, Natasha Favoretto. LANZARINI, Vivian. ONUCHIC, Luiz Fernando. KOCH, Vera Hermina Kalika. Aspectos clínicos da doença renal policística autossômica recessiva DRPAR. Brazilian Journal of Nephrology. 2010;32. p. 263-7.

BETTENCOURT-DIAS, M.. HILDEBRANDT, F.. PELLMAN, D.. WOODS, G.. GODINHO, S. A. Centrosomes and cilia in human disease. Trends Genet. 2011;27(8). p. 307-15.

WALKER, R. V.. KEYNTON, J. L.. GRIMES, D. T.. SREEKUMAR, V.. WILLIAMS, D. J.. ESAPA, C., et al. Ciliary exclusion of Polycystin-2 promotes kidney cystogenesis in an autosomal dominant polycystic kidney disease model. Nat Commun. 2019;10(1). p. 4072.

TOLEDO, Nathalia Bufolin. MAIMONE, Juliana Borges Risolia. MARCOS, Alléxya Affonso Antunes. LEITE, Eduardo Henrique Morizot. COUTO JUNIOR, Abelardo de Souza. Síndrome de Bardet-Biedl: série de caso e revisão da literatura. Revista Brasileira de Oftalmologia. 2018;77. p. 360- 2.



ADAMIOK-OSTROWSKA, A.. PIEKIELKO-WITKOWSKA, A. Ciliary Genes in Renal Cystic Diseases. Cells. 2020;9(4). p.

WALLINGFORD, J. B. Planar cell polarity signaling, cilia and polarized ciliary beating. Curr Opin Cell Biol. 2010;22(5). p. 597-604.

BRASILEIRO FILHO, G. Bogliolo: patologia. 9ª ed ed. Rio de Janeiro: Guanabara Koogan; 2016. 1556 p.

CAVALINI, J. F.. AIELLO, V. D.. EBAID, M. Ausência de Conexão Atrioventricular à Direita. Apresentação Morfológica e Clínica quando o Ventrículo Principal é Morfologicamente Direito. Arq Bras Cardiol. 2020;71(6). p. 793-6.

POPRZECZKO, M.. BICKA, M.. FARAHAT, H.. BAZAN, R.. OSINKA, A.. FABCZAK, H., et al. Rare Human Diseases: Model Organisms in Deciphering the Molecular Basis of Primary Ciliary Dyskinesia. Cells. 2019;8(12). p.

HORNEF, N.. OLBRICH, H.. HORVATH, J.. ZARIWALA, M. A.. FLIEGAUF, M.. LOGES, N. T., et al. DNAH5 mutations are a common cause of primary ciliary dyskinesia with outer dynein arm defects. Am J Respir Crit Care Med. 2006;174(2). p. 120-6.

ORTEGA, Hugo Alejandro Vega. VEGA, Nelson de Araujo. SANTOS, Bruno Quirino dos. MAIA, Guilherme Tavares da Silva. Discinesia ciliar primária: considerações sobre seis casos da síndrome de Kartagener. Jornal Brasileiro de Pneumologia. 2007;33. p. 602-8.

OLM, Mary Anne Kowal. CALDINI, Elia Garcia. MAUAD, Thais. Diagnosis of primary ciliary dyskinesia. Jornal Brasileiro de Pneumologia. 2015;41. p. 251-63.

FERMEIRO, Joana. BANDEIRA, Teresa. LOBO, Luísa. PEREIRA, Luísa. Discinesia ciliar primária revisitada: A propósito de três casos clínicos Revista Portuguesa de Pneumologia. 2010;16. p. 837-47.

GOMES, Juliana de Oliveira. SCURO, Gisele. GREGÓRIO, Carla. LOPES, Renato Delascio. GUIMARÃES, Hélio Penna. LOPES, Antonio Carlos. Síndrome de Kartagener. Relato de Caso. Rev Bras Clin Med. 2008;6. p. 210-2.

GÓMEZ-CORREA, Sandra Viviana. RUÍZ-ÁNGEL, Iván David. SALAZAR-DÍAZ, Luis Carlos. Kartagener syndrome, current data on a classical disease. case report. Case reports. 2018;4. p. 137-44.

PÉREZ CRESPO, Ma. del Rocío. FARIÑAS SALTO, Mercedes. CHACÓN AGUILAR, Rocío. NAVAS CARRETERO, Adriana. SANAVIA MORÁN, Eva. ALBI RODRÍGUEZ, Salomé, et al. Síndrome de Kartagener: diagnóstico neonatal. A propósito de un caso. Arch Argent Pediatr. 2019;117(3). p. 292-6.

GONÇALVES, Luiz Flávio Galvão. SOUTO, Fernanda Maria Silveira. FARO, Fernanda Nascimento. MENDONÇA, Rodrigo de Castro. OLIVEIRA, Joselina Luzia Menezes. SOUSA, Antônio Carlos Sobral. Dextrocardia com situs inversus associada à miocardiopatia não compactada. Arquivos Brasileiros de Cardiologia. 2013;101. p. e33-e6.

YILMAZ, Seher. DEMIRTAS, Abdullah. TOKPINAR, Adem. ACER, Niyazi. Dextrocardia and Situs Inversus Totalis in a Turkish Subject: A Case Report. International Journal of Morphology. 2019;37. p. 900-2.



ALJURE REALES, Vicente de Jesús. ÁLVAREZ GALLEGO, Gloria Camila. ÁVILA ESPITIA, Nasly Consuelo. ARRIETA COLEY, Alexandra. ÁNGEL SUÁREZ, Orlando Germany. Situs inversus totalis: revisión de tema con aproximación a la Genética y reporte de casos. Revista Colombiana de Cardiología. 2017;24(1). p. 40-7.

POSTEMA, Merel C.. CARRION-CASTILLO, Amaia. FISHER, Simon E.. VINGERHOETS, Guy. FRANCKS, Clyde. The genetics of situs inversus without primary ciliary dyskinesia. Scientific Reports. 2020;10(1). p. 3677.

GORT HERNÁNDEZ, Magaly. Situs inversus totalis: presentación de un caso. Revista de Ciencias Médicas de Pinar del Río. 2010;14. p. 250-5.

LAMOTTE, Sandee. CNN News. She lived for 99 years with organs in all the wrong places and never knew it. 2019. p.